



Formulation and Evaluation of Herbal Emulgel Containing Witch Hazel Extract for Anti-Inflammatory Activity

Article History:

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Abstract: Background: Inflammatory skin disorders often require long-term topical therapy, where safety, efficacy, and patient compliance are major concerns. Herbal medicines provide a promising alternative to synthetic agents. Witch Hazel (*Hamamelis virginiana*) is known for its anti-inflammatory, antioxidant, and astringent activities. The present study aimed to formulate and evaluate a herbal emulgel containing Witch Hazel extract for anti-inflammatory activity. The bark extract was obtained using hydroalcoholic maceration and incorporated into six different emulgel batches using Carbopol 940, HPMC, and Xanthan gum as gelling agents. Formulations were evaluated for organoleptic properties, pH, viscosity, spreadability, drug content, in vitro diffusion, anti-inflammatory activity by HRBC membrane stabilization, and stability studies. Among the six batches, F5 showed optimal performance with pH 5.9 viscosity 8800 cP, drug content 91%, sustained release up to 83.4% over 13 hours, and 73.78% inhibition in anti-inflammatory assay compared to 93.45% by aspirin. Stability studies revealed no significant changes in physicochemical parameters over 3 months. The findings suggest that Witch Hazel emulgel is a stable, effective, and patient-friendly formulation with potential use for inflammatory skin disorders.

Keywords: Emulgel, Witch Hazel, Anti-inflammatory, Herbal formulation, Topical drug delivery.

INTRODUCTION

Skin is the largest organ of the body and a primary barrier against microbial invasion, injury, and environmental exposure¹. Inflammatory skin disorders such as atopic dermatitis, psoriasis, and seborrheic dermatitis significantly impair quality of life due to chronic itching, scaling, redness, and swelling². Current therapies include corticosteroids, immunosuppressants, and topical creams, but they are often associated with side effects, irritation, and poor

compliance.

Emulgels, a hybrid of emulsion and gel systems, combine the advantages of both dosage forms, providing better stability, cosmetic acceptability, non-greasy texture, and sustained drug release³. They are suitable for localized drug delivery, bypass first-pass metabolism, and improve patient compliance. Herbal drugs, when incorporated in emulgels, offer enhanced therapeutic efficacy with reduced risk of side effects⁴.

Witch Hazel (*Hamamelis virginiana*) is a well-known medicinal plant with phytoconstituents such as tannins, flavonoids, and gallic acid, responsible for its anti-inflammatory, antimicrobial, and antioxidant activities⁵. It has been traditionally used in treating skin irritations, wounds, eczema, and hemorrhoids⁶. However, conventional formulations face limitations like poor penetration and instability. Thus, developing a Witch Hazel-based herbal emulgel provides a modern, stable, and effective topical delivery system⁷.

The present study focuses on the formulation and evaluation of herbal emulgel containing Witch Hazel extract for anti-inflammatory activity.

MATERIALS AND METHODS

Materials

Witch hazel bark extract, Carbopol 940, HPMC, Xanthan gum, Span 20, Tween 80, Light liquid paraffin, Methyl paragon, propylene glycol,

triethanolamine, distilled water.

Methods

Extraction of Witch Hazel

1. Bark was shade-dried and coarsely powdered.
2. Macerated with hydroalcoholic solvent (70:30 ethanol-water) for 72 hours at room temperature with occasional shaking.
3. Filtrate was concentrated under reduced pressure at <40°C using a rotary evaporator.
4. Extract was stored in amber-colored bottles for further use.

Method of Preparation of Emulgel

Step 1: Oil-in-water emulsion prepared by heating oil phase and aqueous phase separately at 60–70°C, then combining with continuous stirring.

Step 2: Gel bases prepared individually with Carbopol 940, HPMC, and Xanthan gum.

Step 3: Emulsion incorporated into gel base in 1:1 ratio with gentle stirring to avoid air entrapment.

Table No. 1: Formulation table

Composition	F1	F2	F3	F4	F5	F6
Witch Hazel Extract (gm)	1	1	1	1	1	1
Carbopol 940 (gm)	1	1.5	-	-	-	-
HPMC (gm)	-	-	1	1.5	-	-
Xanthan Gum (gm)	-	-	-	-	1	1.5
Propylene Glycol (ml)	5	5	5	5	5	5
Methyl Paraben (gm)	0.03	0.03	0.03	0.03	0.03	0.03
Propyl Paraben (gm)	0.01	0.01	0.01	0.01	0.01	0.01
Light liquid paraffin (ml)	7	7	7	7	7	7
Span 20 (ml)	1	1	1	1	1	1
Tween 80 (ml)	0.5	0.5	0.5	0.5	0.5	0.5
Triethanolamine (ml)	1.2	1.2	-	-	-	-
Distilled water	qs	qs	qs	qs	qs	qs

Evaluation of Witch Hazel Emulgel

1. Physical Appearance:

The formulated emulgel was examined visually to assess its colour, uniformity, consistency, and any sign of phase separation. The colour contrast was checked against both white and black backgrounds. To evaluate consistency, a small amount was applied to the skin and observed.

2. pH Measurement:

Measuring the pH is essential for ensuring the emulgel is suitable for topical application. An ideal pH range between 5.5 and 6.0 is preferred to match the skin's natural pH and avoid irritation. The pH was determined using a calibrated digital pH meter by immersing the glass electrode into the sample. Each formulation was tested three times, and the mean value was recorded.

3. Viscosity:

Viscosity was determined using a Brookfield Viscometer. A measured quantity of the emulgel was placed in a beaker and allowed to settle for 30 minutes at room temperature (25–30°C). The spindle (spindle no.6) was carefully positioned to avoid contact with the bottom of the container and rotated at 100 RPM for 10 minutes. The viscosity reading was then recorded.

4. Spreadability:

Spreadability was tested using a setup involving two glass slides and a wooden block with a pulley mechanism. A fixed amount (1 g) of emulgel was placed between two slides, with one slide mounted on a block. A standard weight (50 g) was applied, and the time taken for the upper slide to move was noted. The spreadability was calculated using the formula:

$$S = ML/T$$

Where:

S = Spreadability

M = Weight attached to the upper slide (g)

L = Distance moved by the slide (cm)

T = Time taken to separate the slides (sec)

5. Drug Content Estimation:

To determine the active ingredient concentration, 1 g of emulgel was dissolved in 100 ml of methanol. After proper dilution and filtration through a 0.45 µm membrane, the solution's absorbance was measured at 271 nm using a UV-Visible spectrophotometer.

6. In Vitro Drug Diffusion Studies:

The diffusion study was performed using a modified

dissolution basket-type apparatus with a two-sided open glass cylinder. A dialysis membrane (HiMedia, MW 12–14 kDa) was fixed at one end, and 1 g of the formulation was placed inside. The setup was immersed in 200 mL of phosphate buffer (pH 7.2) maintained at 37±1°C. At predetermined intervals (1 to 13 hours), 0.5 mL samples were withdrawn and replaced with fresh buffer (pH 6.8) to maintain sink conditions. Samples were analyzed at 271 nm using a UV-visible spectrophotometer, and % cumulative drug release (%CDR) was calculated.

7. Evaluation of Anti-Inflammatory Activity – Human Membrane Stabilization Assay (HRBC Method) To assess the anti-inflammatory activity of the prepared witch hazel emulgel, the human red blood cell (HRBC) membrane stabilization method was employed. Blood samples were obtained from a healthy volunteer who had not consumed any non-steroidal anti-inflammatory drugs (NSAIDs) for at least two weeks prior to collection. The blood was collected into sterile centrifuge tubes and centrifuged at 3000 rpm for 10 minutes to separate the red blood cells. The sedimented RBCs were washed three times with an equal volume of normal saline (0.9% NaCl) to remove any plasma and other components. After the final wash, the packed cell volume was noted, and the erythrocytes were re-suspended in normal saline to prepare a 10% v/v RBC suspension. This suspension was then used in further steps to evaluate the membrane stabilizing effect of the test emulgel, which indirectly reflects its potential Anti-inflammatory properties.

RESULT AND DISCUSSION

Organoleptic properties

The drug was studied for their organoleptic properties like colour, odour, taste, crystallinity and Ph observation was recorded in table 1.

Table no 2: organoleptic Properties of Witch Hazel

Parameters	Result
Colour	Light Brown
Odour	Mild
Appearance	Fine to Moderate Coarse powder

Solubility Study

The solubility of witch hazel is found to be as per following table

Table no 3: Solubility of Witch Hazel

Sr. No	Solvent System	Result
1	Ethanol	Soluble
2	Methanol	Soluble
3	Water	Soluble
4	Glycerine	Partially soluble
5	Propylene glycol	Soluble

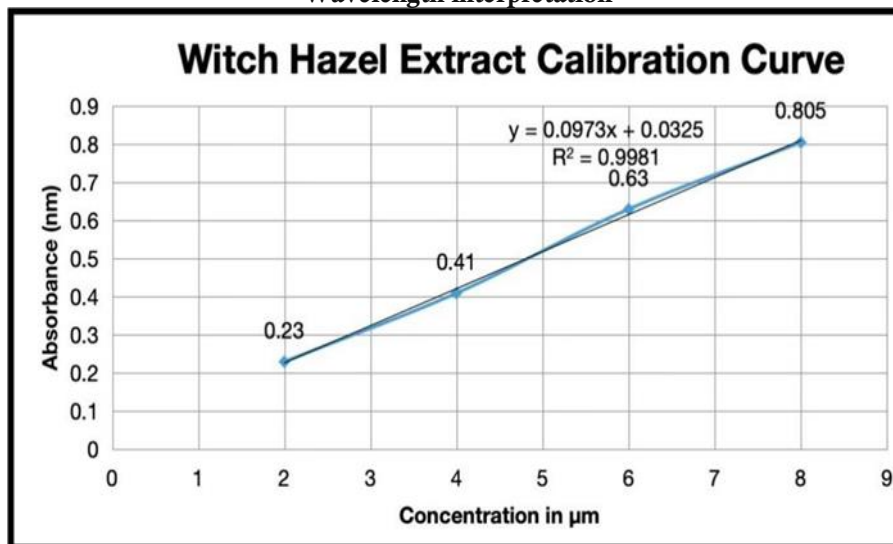
Phytochemical Screening

Preliminary qualitative phytochemical screening revealed the presence of the following :

Table No 4: Phytochemical tests

Phytoconstituent	Test Performed	Observation	Inference
Tannins	Ferric Chloride test-Lead Acetate test	Blue black or green colour	Present
Flavonoids	Shinoda Test Alkaline Reagent Test	Magenta Colour	Present
Phenolic compounds	Ferric Chloride test	Deep Blue black or green colour	Present
Saponins	Foam Test	Persistent froth formation	Present
Terpenoids	Salkowski Test	Raddish brown	Present

Wavelength interpretation



Concentration (µg/ml)	Absorbance 271nm
2	0.230
4	0.410
6	0.630
8	0.805

FTIR Spectrum Interpretation

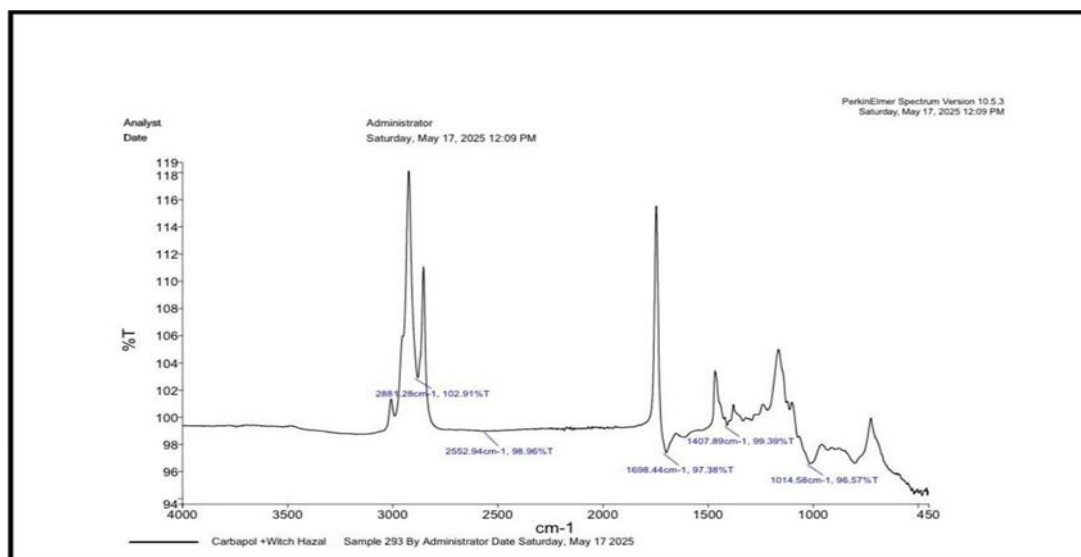
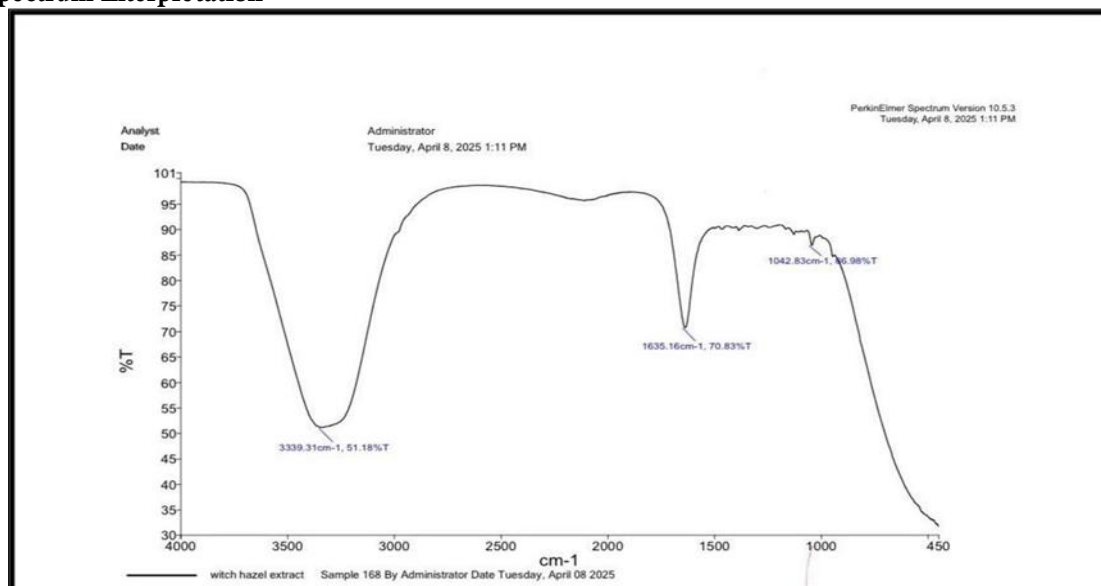


Fig No. 2: FTIR of Witch hazel extract
Fig No. 2: FTIR of Witch hazel extract + Carbopol 940

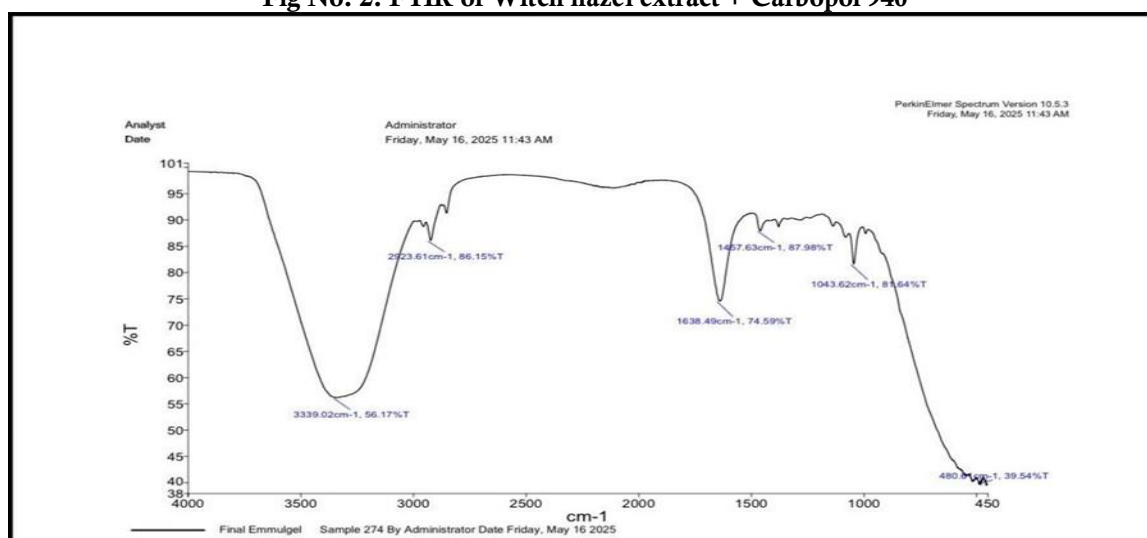
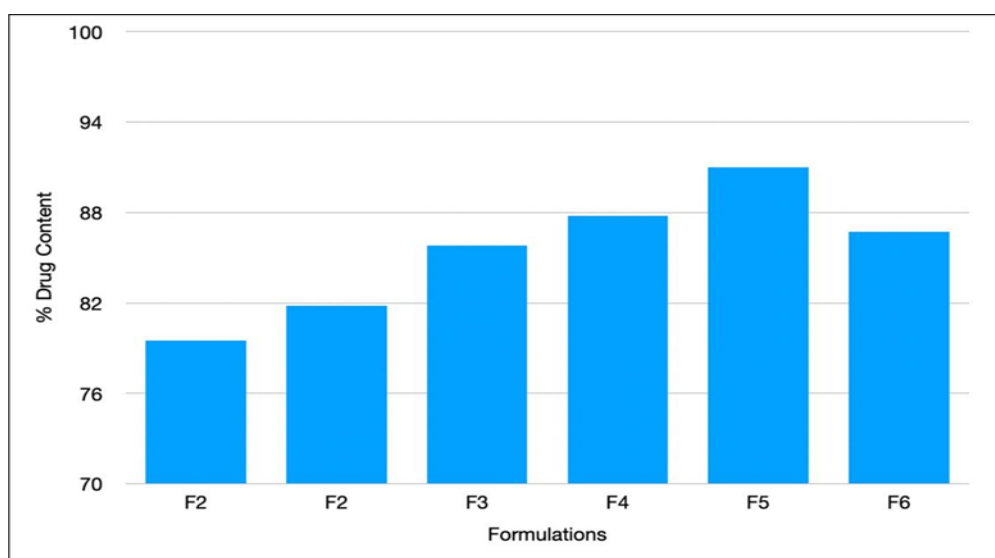


Fig No. 3: FTIR of Witch hazel extract Emulgel Formulation

Evaluation of Emulgel

Table No 5: Evaluation parameters of emulgel

Formulation code	Appearance	Viscosity (CPS)	pH	Spreadability (gm/sec)
F1	Semi-transparent, brownish	5800	5.9	9.7
F2	Semi-transparent, brownish	7200	5.8	8.3
F3	Semi transparent, brownish	4300	5.5	9.7
F4	Semi transparent, brownish	6200	5.7	9.2
F5	Semi transparent, brownish	8800	5.9	7.7
F6	Semi transparent, brownish	8300	6.0	7.9



Drug Content

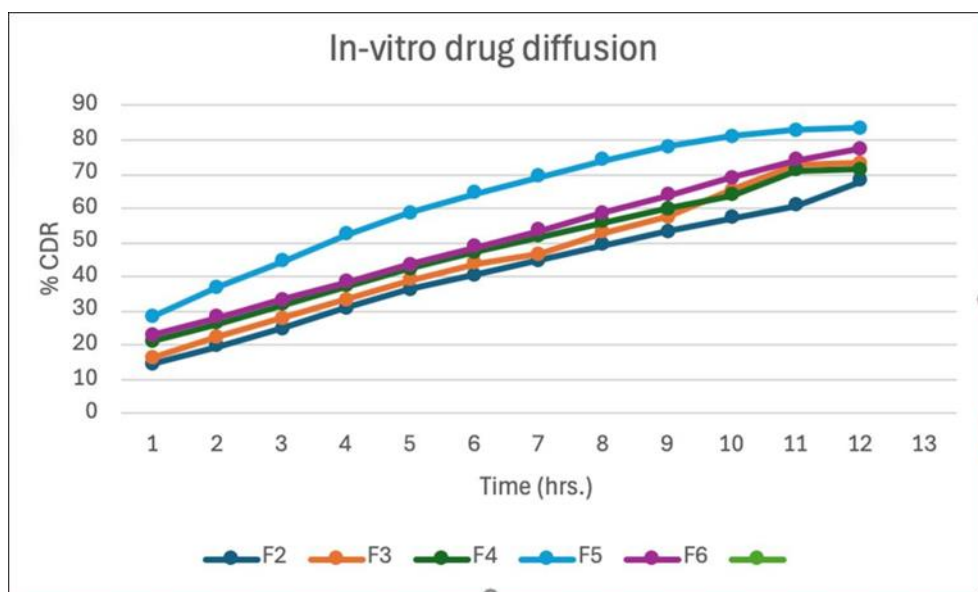
Table No 6: Drug Content

Batches	F1	F2	F3	F4	F5	F6
% Drug Content	79.5	81.8	85.8	87.8	91.0	86.7

In Vitro Drug Diffusion Study

Table No 7: In vitro drug diffusion study

Time (hrs.)	F1 (%)	F2 (%)	F3 (%)	F4 (%)	F5 (%)	F6 (%)
1	15.2	14.7	12.5	15.5	20.0	18.0
2	21.0	19.8	16.3	21.2	28.5	23.1
3	26.5	25.0	22.5	26.3	36.8	28.2
4	32.0	30.8	27.9	31.8	44.6	33.3
5	37.6	36.3	33.4	37.3	52.3	38.4
6	42.0	40.6	39.0	42.5	58.7	43.5
7	46.5	44.8	43.8	47.3	64.5	48.6
8	50.8	49.2	46.5	51.8	69.2	53.7
9	54.9	53.4	53.1	56.0	74.1	58.8
10	58.7	57.2	57.6	59.9	78.0	63.9
11	62.4	60.9	65.3	63.7	81.2	69.0
12	69.5	68.2	73.0	71.0	83.0	74.1
13	69.7	68.8	73.2	71.3	83.4	77.4



In Vitro Anti- Inflammatory Assay by HRBC Membrane Stabilization Method

Fresh human blood (10 ml) will be collected in heparinized centrifuge tubes and centrifuged at 3000 rpm for 10 min and washed 3× with an equal volume of normal saline solution. The volume of the blood will be measured and reconstituted as a 10% v/v suspension with normal saline. The reaction mixture consisted of formulation of 1 ml and 1 ml of 10% red blood cell suspension. For the control, saline will be added. Aspirin will be used as a standard drug (positive control). The samples will be incubated at 56 °C for 30 min, centrifuged at 2500 rpm for 5 min and the absorbance of the supernatant measured at 560 nm

Table No 7: In vitro Anti-Inflammatory Assay by HRBC

Sample Code	Concentration (ul/ml)	Test 1	Test 2	Test 3	Mean	% Hemolysis	% Stabilisation
Control		1.23	1.23	1.22	1.22	-	-
Standard (Aspirin)	20	0.35	0.36	0.36	0.36	29.5%	70.5%
	40	0.29	0.29	0.29	0.29	23.77%	76.23%
	60	0.15	0.15	0.15	0.15	12.29%	87.8%
	80	0.13	0.13	0.13	0.13	10.65%	89.35%
	100	0.08	0.08	0.08	0.08	6.55%	93.45%
F 5 (Witch hazel)	20	0.92	0.90	0.89	0.90	73.77%	26.23%
	40	0.74	0.72	0.71	0.72	59.01%	40.99%
	60	0.58	0.60	0.56	0.58	47.54%	52.46%
	80	0.42	0.40	0.43	0.41	33.6%	66.4%
	100	0.34	0.31	0.33	0.32	26.22%	73.78%

The F5 at concentration range 20µg/mL to 100 µg/mL may protects the human erythrocyte membranes against lysis induced by hypotonic solution. At concentration of 100µg/mL, F5 inhibited of 73.78 % RBC haemolysis as compared with 93.45% produced by Aspirin at 100µg/mL. Since human red blood cell membranes are similar to lysosomal membrane components, the prevention of hypotonicity induced HRBC membrane lysis was taken as a measure of anti-inflammatory activity of drugs.

Stability Study

The formulated emulgel was filled into 5 g aluminum collapsible tubes and subjected to accelerated stability testing under controlled conditions of 40 ± 2°C temperature and 75 ± 5% relative humidity for a period of three month as per (ICHQ1A(R2) guideline.

Table No 7: Stability study

Month	Physical Appearance	pH	Viscosity (Cps)	Drug Content %	Drug Diffusion %
Initial	Slightly-opaque, smooth	5.9	8800	91.0	83.4
1 Month	No change	5.8	8750	90.9	83.2
2 Months	No change	5.6	8700	90.7	83.0
3 Months	No change	5.5	8660	90.5	82.9

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